# **Matthew McGowan**

Molecular Plant Science Washington State University Pullman, WA 99163

## **Education**

Ph.D.	Molecular Plant Science	Washington State University	2020 (expected)
M.S.	Biomedical Science	Kansas State University	2014
B.S.	Microbiology, Anthropology	Kansas State University	2011

Phone: (913)-484-9395

email: matt.mcgowan@wsu.edu

# **Academic Experience**

Graduate Research Assistant. Department of Crop and Soils, Washington State University. Current.

- Incorporates conditional co-expression network data into GWAS for ensemble analysis
- Develops, maintains, and troubleshoots computational tools for high-throughput genotyping and RNA-seq network analysis using R, Python, and Slurm Workload Manager
- Leverages high-performance computing clusters to analyze complex gene co-expression networks
- Performs QC analysis on publicly available transcriptomic data for RNA-seq network research
- Maintains internal Github project repositories for research reproducibility
- Completed coursework in bioinformatics, statistical genomics, and data science
- Presented biological research at international scientific conferences

Graduate Research Assistant. Department of Pathobiology, Kansas State University. Mar 2010 – Dec 2013.

- Deployed epidemiologically sound strategies to isolate representative commensal bacterial populations and extract nucleic acids from animal blood, feces, and tissue for further phenotypic and genotypic analysis
- Contributed scientific research that resulted in co-authorship on a peer-reviewed published paper
- Utilized PCR and real-time PCR approaches to measure the burden of antibiotic resistance present in cattle herds
- Introduced high-throughput automated equipment into laboratory processes to decrease data turnaround time, improve consistency, and minimize labor requirements
- Provided USDA and Canadian resistance surveillance programs with Sensititre MIC and growth curve assay data for foodborne pathogen strains
- Collaborated on a large multi-institutional grant project aimed at developing antibiotic resistance intervention strategies by presenting research, editing papers, and acting as conference rapporteur to assemble annual reports on project milestones
- Completed coursework required to complete a M.S. in Biomedical Science

### **Non-Academic Experience**

Research Associate. Heartland Plant Innovations. Manhattan, KS. Jan 2014 – Dec 2015.

- Coordinated daily and long-term operations of a molecular biology services laboratory
- Managed corporate genotype-by-sequencing projects for diverse plant species: wheat, barley, corn, soybean, and strawberry
- Streamlined automated molecular methodologies to reduce turnaround time and material costs
- Maintained laboratory equipment, wrote standard operating protocols, and trained laboratory technicians in proper equipment usage
- Curated project databases and handled quarterly data backups
- Assisted in seasonal wheat doubled haploid production processes including mother plant emasculation, pollination, embryo rescue, tissue culture, colchicine treatment, and greenhouse management
- Communicated with customers to assess project goals, deadlines, and progress updates

#### **Academic Advisors**

Dr. Stephen Ficklin (2017 – present) Dr. Zhiwu Zhang (2016 – present)

Dr. H. Morgan Scott (2012-2014)

# **Publications and Scholarly Work**

### **Publications**

- M. Mcgowan, "Calling SNPs Out of DNA Array for Diverse Hexaploid Wheat Population by Using Gaussian Mixture Models", Proceedings of the World Congress on Genetics Applied to Livestock Production, vol. Technologies - Genotyping, p. 946, 2018.
- Kanwar et al, "Effects of Ceftiofur and Chlortetracycline Treatment Strategies on Antimicrobial Susceptibility and on tet(A), tet(B), and blaCMY-2 Resistance Genes among E. coli Isolated from the Feces of Feedlot Cattle," PLoS ONE 8(11)" e80575, doi: 10.1371/journal.pone.0080575

#### **Theses**

• McGowan, M., *The Epidemiology of Tetracycline and Ceftiofur Resistance in Commensal Escherichia coli*, M.S. Thesis, Kansas State University, Department of Pathobiology, 2014.

#### Presentations at Conferences, Symposia, and Workshops

- Conditional Gene Co-Expression Networks for Aphanomyces Root Rot Response (ARR) in Lentil, Plant and Animal Genome Conference, San Diego, CA (January 2019)
- Improved SNP Calling for Diverse Hexaploid Wheat Using GGcall, World Congress on Genetics Applied to Livestock Production, Auckland, NZ, (February 2018)
- Calling SNPs Out of DNA Array for Diverse Hexaploid Wheat Population by Using Gaussian Mixture Models, Plant and Animal Genome conference, San Diego, CA (January 2018)
- SNPs from Sequence: Genotyping in Wheat, Western Wheat Workers Meeting, Pullman, WA (July 2016)
- Relationships between antibiotic use and resistance genetics in commensal E. coli using pairwise bacterial growth parameters, Federation of European Microbiological Society (FEMS) general conference, Leipzig, Germany, (July 2013)
- Genotypic surveillance of tetracycline resistance in Canadian beef cattle 2002-2011, Antimicrobial Resistance in Animals and the Environment (ARAE) Symposium, Ghent, Belgium, (July 2013)
- Antibiotic use versus antibiotic resistance profiles of commensal E. coli in beef cattle: explaining their association via bacterial growth parameters, Conference of Research Workers in Animal Diseases (CRWAD), Chicago, IL, (December 2012)

# **Academic Awards**

- Howard Hughes Undergraduate Research Scholar, 2006-2008
- WSU Hardware Engineering Hackathon 2<sup>nd</sup> Place, Cyberthumb plant sensor, 2017

### Institutional, Professional, and Outreach Activities

- Board Member of the College of Veterinary Medicine Graduate Research Association, 2012-2013
- Molecular Plant Science Seminar Organizing Committee Member, 2016-2018
- Franklin Elementary School Science Fair Judge, 2017-2019
- Pullman USDA ARS Peoples' Garden Volunteer, 2017-2018